

area of approximately 1.5 m<sup>2</sup> (assuming an elliptical shape) and is around 150 m from the nearest stream, while wallow #2 has an approximate area of 3.4 m<sup>2</sup> and is about 50 m away from the nearest stream. In September 2012 we revisited both wallows and found a subadult female *A. oxylophus* (SVL: 56.1 mm) in standing water at wallow #2. Wallow #1 no longer contained water.

Historically, *A. oxylophus* has been observed living in and very close to streams, and has not been observed beyond the riparian zone (Savage 2002, *op. cit.*; Vitt et al 1995. *Can. J. Zool.* 73:1918–1927). To our knowledge, our observations are the first of *A. oxylophus* using peccary wallows. Our observations suggest that these tropical anoles are able to utilize a wider range of habitats than has previously been reported, which may have important implications for conservation efforts. In the Amazon, White-lipped Peccaries (*Tayassu pecari*) have been recognized as ecosystem engineers because of their habit of maintaining wallows, which amphibians use as reproductive habitat (Beck et al. 2010. *J. Trop. Ecol.* 26:407–414; Zimmerman and Bierregaard 1986. *J. Biogeog.* 13:133–143). Some Central American amphibian species have recently been observed using Collared Peccary wallows for breeding (Reider and Ream, ms. in review). The presence of *A. oxylophus* in and around Collared Peccary wallows indicates that wallows might be important, albeit temporary, habitat for aquatic reptiles in addition to amphibians. Our observations might imply that wallows serve as important stops for *A. oxylophus* moving between streams and warrant additional research to investigate the dispersal patterns of this species. Additionally, future studies of the biology of *A. oxylophus* should consider that some individuals might be spending a portion of their time away from streams.

We thank the staff at La Selva Biological Station for logistical support. These observations were made possible by funds from the Organization for Tropical Studies and the National Science Foundation Research Experience for Undergraduates program.

**KELSEY A. REAM**, Departments of Biological and Environmental Sciences, Allegheny College, 520 North Main St., Meadville, Pennsylvania 16335, USA (e-mail: reamk@allegheny.edu); **KELSEY E. REIDER**, Department of Biological Sciences OE167, Florida International University, 11200 SW 8<sup>th</sup> St., Miami, Florida 33199, USA (e-mail: reider.12@gmail.com).

**ANOLIS SAGREI (Brown Anole). NEST-SITE CHOICE.** *Anolis sagrei* is native to Cuba and the Bahamas, and has been introduced throughout many tropical and subtropical regions of the world (Kolbe et al. 2004. *Nature* 431:177–181). Although *A. sagrei* has served as a model for many ecological and behavioral studies (Losos 2009. *Lizards in an Evolutionary Tree: Ecology and Adaptive Radiation of Anoles*. Univ. of California Press. 528 pp.), relatively little is known about its oviposition behavior. Like all anoles, female *A. sagrei* produce a single egg clutch, for the duration of a breeding season lasting from April to October (Lee et al. 1989. *Copeia* 1989:930–937). Despite their frequent egg production, the lack of published information about their oviposition sites is largely attributable to difficulty in finding nests.

During field work from 29 September to 9 October 2012, we documented five *Anolis* eggs on two spoil islands in the intra-coastal waterway near Marineland, Florida, USA (29.640042°N, 81.214419°W; WGS 84). All eggs were found in soil beneath wooden cover objects. On two occasions, two eggs were found under the same cover object (four eggs total). The fifth egg was found under a board that also contained hundreds of individuals of an unknown ant species. Two of the eggs (found under the same object) were empty shells that hatched earlier, desiccated,

or were depredated; the other three eggs were intact and turgid (including the egg in the presence of ants). However, one of the intact eggs had disappeared less than one hour after discovery, possibly depredated upon by a fiddler crab at the study site (*Uca* sp.). Given the high densities of *A. sagrei*, it is unlikely that these eggs belonged to another lizard species (e.g., *Anolis carolinensis*, the only other lizard on these islands, which occurs at relatively low densities).

To assess microhabitat characteristics of the oviposition sites, we took soil samples to measure moisture content and hemispherical photos to measure canopy openness (methodology of Doody et al. 2006. *Herpetol. Rev.* 37:49–52) at each oviposition site. The three intact eggs were in soil that had an average moisture content of 15.8% (range 15.0–16.5%), whereas the soil moisture content around the non-intact eggs was 63.0%. The intact eggs were found at relatively open sites (mean canopy openness = 46.7%, range: 43.7–51.2%) compared to the site with the non-intact eggs (14.8% canopy openness). Canopy openness from these five nests (mean = 33.8%) did not differ from canopy openness measured at 40 randomly selected locations on the islands (mean=34.9%), suggesting that female *A. sagrei* choose oviposition sites randomly with respect to canopy cover.

Although nest-site information has been reported for other *Anolis* species (Andrews 1988. *Oecologia* 76:376–382; Rand 1967. *Herpetologica* 23:227–230), we are not aware of previously published information for *A. sagrei* in the field. Laboratory-based studies have shown that females select relatively moist conditions that enhance egg hatching success and offspring survival (Reedy et al. 2013. *Behav. Ecol.* 24:39–46). Although we do not know what moisture conditions were present at the time of oviposition for the eggs reported here, we nonetheless provide some baseline data on moisture levels that eggs experience in the field. Our small sample suggests that female nest-site choice is not based on canopy cover—a finding that contrasts with other studies of reptiles (e.g., Angilletta et al. 2009. *Ecology* 90:2933–2939; Warner and Shine 2008. *Anim. Behav.* 75:861–870), and hence warrants focused studies on nest-site choice and egg ecology in *Anolis sagrei*.

**DAVID M. DELANEY**, Department of Biological Sciences, Eastern Illinois University, Charleston, Illinois 61920, USA (e-mail: dmdelaney3@eiu.edu); **AARON M. REEDY**, Department of Biology, University of Virginia, Charlottesville, Virginia 22904, USA; **TIMOTHY S. MITCHELL**, Department of Ecology, Evolution, and Organismal Biology, Iowa State University, Ames, Iowa 50011, USA; **ANDREW M. DURSO**, Department of Biology, Utah State University, Logan, Utah 84321, USA; **KEVIN P. DURSO** and **AL-EXANDRA J. MORRISON**, College of Natural Resources, North Carolina State University, Raleigh, North Carolina 27695, USA; **DANIEL A. WARNER**, Department of Biology, University of Alabama at Birmingham, Birmingham, Alabama 35294, USA.

**ASPIDOSCELIS LAREDOENSIS × ASPIDOSCELIS GULARIS (Laredo Striped Whiptail × Texas Spotted Whiptail). HYBRID GYNANDROMORPH.** We evaluate the identity and reproductive status of a whiptail lizard from a site of syntopy involving diploid hybrid-derived parthenogenetic *Aspidoscelis laredoensis* and its maternal progenitor *A. gularis* (Bickham et al. 1976. *Herpetologica* 32:395–399; McKinney et al. 1973. *Herpetologica* 29:361–366; Wright et al. 1983. *Herpetologica* 39:410–416) situated in Texas (Walker 1987. *Texas J. Sci.* 39:313–334). The specimen is unlike any other in color pattern among a combined total of >1000 specimens of these species examined by us between 1984–2009 from México and Texas. Consequently, it was hypothesized to be